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ARTICLE DETAILS

Article history:

Received 11 April 2020

Accepted 26 April 2020

Available online 02 May 2020

Keywords:

Silver Nanoparticles

Biological Green Synthesis

Colpomenia sinuosa

ABSTRACT

In the present study energy efficient, economically scalable colloidal silver (Ag) nanoparticles were biosynthesized from marine brown seaweed *Colpomenia sinuosa* by green synthesis method. The marine macroscopic brown seaweed *Colpomenia sinuosa* was used in the experimental study for the biosynthesis of silver nanoparticle since they are rich in phytochemicals, bioactive compounds and secondary metabolites which has reducing agents that may be environmentally acceptable and eco-friendly. The biosynthesized silver nanoparticles from marine macroscopic brown seaweed were characterized by UV-vis spectroscopy which confirmed the surface plasmon resonance of silver nanoparticles, Fourier transform infrared (FT-IR) spectroscopy to identify the presence of various functional groups in biomolecules responsible for the bio reduction of Ag⁺ and capping/stabilization of silver nanoparticles. X-ray diffraction (XRD) to observe face center cubic (fcc) and crystalline nature of silver nanoparticles, thermogravimetric analysis (TGA) which revealed the thermal stability and purity of the silver nanoparticles. Particle size distribution and morphology were investigated by scanning electron microscope (SEM) which showed silver nanoparticles in the size range of 54-85 nm. The particle distribution under different nanometers was analyzed using transmission electron microscopy (TEM).

1. Introduction

Nanotechnology involves the characterization, fabrication of structures, materials or particles that have at least 1–100 nm in length. The field of nanotechnology is one of the most active research areas in modern materials science. Nanoparticle research is an ever-increasing scientific research interest, especially over the last couple of decades. The most effectively studied nanoparticles today are those made from noble metals, in particular Ag, Pt, Au and Pd. Nanoparticles and nanomaterials are quickly becoming part of everyday life. In the current scenario, the use of nanoparticles in biomedical applications such as drug delivery [1-3], cancer-cell diagnostics [4-7] and therapeutics [8] has given nanotechnology a new dimension. Biomolecules have been used for nanomaterial synthesis/functionalization and in subsequent applications for decades [9]. Nanoscience involves investigation into learning new properties of nano size, materials [10]. Nanotechnology is also being utilized in medicine for diagnosis, therapeutic drug delivery and the development of treatments for many diseases and disorders [11]. Silver nanoparticles are one of the promising products in the Nanotechnology industry. The development of consistent processes for the synthesis of silver nanomaterials is an important aspect of current nanotechnology research. One such promising process is the green synthesis. Silver nanoparticles can be synthesized by several physical, chemical and biological methods [12]. A possible application of silver nanoparticles is its potential as a catalyst. Jiang et al. [13] have proved the catalytic potential of nanosilver spheres. Nanosilver (silver nanoparticle, materials) have a wide range of applications including spectrally selective coating for solar energy absorption [14, 15], catalysis in chemical reactions [16], surface-enhanced Raman scattering for imaging [17], and antimicrobial sterilization [18-20]. Because of their effective antimicrobial properties and low toxicity to mammalian cells, silver nanoparticles have become one of the most commonly used nanomaterials in consumer products [21, 22]. The scientific and practical interest in silver nanoparticles was exclusively caused by the possibility of their use as highly dispersed supports for

enhancing the signals from organic molecules in the Raman spectroscopy [23]. A synergistic combination of nanotechnology and biotechnology provides unprecedented opportunities for addressing many of the current challenges in cancer diagnosis and therapy [24-28]. The literature survey revealed that the nanoparticle synthesis using biological sources like algae/seaweeds has been unexplored and unexploited [29-33]. In the present study the green synthesis of silver nanoparticles from marine macroscopic brown seaweed *Colpomenia sinuosa* (Mertens ex Roth) Derbes and Solier was biosynthesized and their characterization was studied by using UV-vis spectroscopy, Fourier transform infrared (FT-IR) spectroscopy, X-ray diffraction (XRD), thermogravimetric analysis, scanning electron microscope (SEM) and transmission electron microscopy (TEM).

2. Experimental Methods

2.1 Collection and Preparation of Seaweed Extract

The marine brown seaweed *Colpomenia sinuosa* (Mertens ex Roth) Derbes and Solier was collected from the intertidal regions of Leepuram, Kanyakumari District (Latitude 8°14'23.10" N, Longitude 77°20'04.02" E); South East Coast of Tamilnadu during summer. Collected seaweed was washed with sea water for eliminating impurities such as sand, rocks, epiphytes and epifauna. The washed samples were preserved with 5 -10% formaldehyde in sea water and transported to the laboratory in a box containing slush ice. The fumes of the formaldehyde would help to fix and preserve the seaweed material. In the laboratory, the samples were washed thoroughly in running tap water to remove salt and washed three times using distilled water which may remove metallic compounds and it was shade dried at room temperature (37 °C) for 10 days. The dried seaweed materials were crushed by using mortar and pestle to get the powder form and it was stored in an air-tight container. About 1 g of crushed seaweed powder was added with 100 mL of distilled water in 250 mL conical flask and boiled for 5-10 minutes at 60-80 °C. The crude extract was collected and stored at 4 °C for experimental use [34].

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2.2 Biosynthesis of Silver Nanoparticles

The crude extract of the experimental marine brown seaweed *Colpomenia sinuosa* (Mertens ex Roth) Derbes and Solier was used for the synthesis of silver nanoparticles. Silver nitrate (AgNO_3) (SD fine) was used for the synthesis of silver nanoparticles and double-distilled, deionized water was used for all the experiments. The silver nanoparticle formation was carried out by taking 500 mg of dry, shade dried powder samples of *Colpomenia sinuosa* in a 250 mL Erlenmeyer flask with 10^{-3} M aqueous AgNO_3 solution and was incubated at room temperature. The pH was checked during the course of reaction, and it was found to be 5.09. Nearly 95% of bio reduction of AgNO_3 ions occurred within 24 hr at stirring condition. The biosynthesis of silver nanoparticles was characterized by UV Vis spectroscopy; size and morphology by employing SEM and TEM, structure from X-ray diffraction (XRD) technique, stability of silver nanoparticles from Thermo gravimetric analysis (TGA) and biomolecules involved in the capping agent of silver nanoparticles from Fourier transform infrared (FT-IR) spectroscopy.

3. Results and Discussion

3.1 Visual Examination

The biosynthesis of silver nanoparticles was primarily identified by color change during exposure of crude seaweed extract of *Colpomenia sinuosa* into aqueous solution of silver ions is shown in Fig. 1. The shade dried powder preparations of the experimental seaweed *Colpomenia sinuosa* were added in 10^{-3} M silver nitrate solution and allowed to react at 121°C for 20 minutes. The colour of the reaction solution changed to dark reddish brown. The control (without seaweed powder) showed no colour formation. Formation of the colour arises due to the excitation of surface plasmon vibrations where the metabolites in the seaweed extract act as the capping agent. The colour of the solution gradually intensified on heating which clearly indicates and confirms the formation of silver nanoparticles. After 24 hr, there is no significant color change, indicating the saturation of the reaction of silver nanoparticle formation.



Fig. 1 Aqueous extract of *Colpomenia sinuosa* before and after synthesis of silver nanoparticles

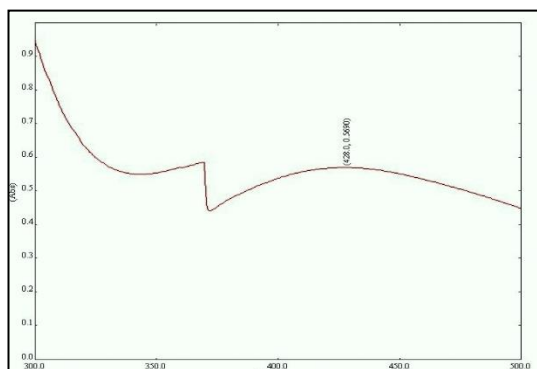


Fig. 2 UV Visible spectral analysis of silver nanoparticles bio-synthesized from *Colpomenia sinuosa*

3.2 UV-Visible Spectroscopic Analysis

The silver nanoparticles synthesized by marine brown seaweed *Colpomenia sinuosa* were analyzed by using UV-Vis spectrophotometer (Labtron LUS-B16). The absorption spectra of silver nanoparticles formed in the reaction solution were characteristic for the seaweed and had specific absorption maxima 428 nm is shown in Fig. 2. The broadening of the peak indicated that the silver nano particles synthesized from the experimental seaweed are polydispersed in nature and the intensity of the band increased with the increase in reaction time. The frequency and

width of the surface plasmon absorption depends largely on the size and shape of the metal nanoparticles as well as on the dielectric constant of the metal and the surrounding medium [35-37]. The interaction with the biomolecules presents in the aqueous part of the reaction solution by the biosynthesized silver nanoparticles from experimental seaweed has been indicated by UV-Visible spectroscopic analysis. There were no little signs of aggregation with the biosynthesized silver nanoparticles solution which were stable for more than six months of observation.

3.3 FT-IR Spectroscopic Analysis (FTIR)

The FTIR spectral measurements were carried out to identify the potential biomolecules in the crude extract of the seaweed *Colpomenia sinuosa* which is responsible for reducing and capping the bio reduced silver nanoparticles. Silver nanoparticles biosynthesized from experimental seaweed *Colpomenia sinuosa* were analyzed using FT-IR spectroscopy is shown in Fig. 3 and Table 1. The local molecular environment of the organic molecules on the surface of the nanoparticles was determined by the IR spectra. Fourier transform infrared spectroscopy (FTIR) is a technique which is used to analyze the chemical composition of many organic chemicals, semiconductor materials, gases, biological samples, inorganics, and minerals. FTIR analysis can give not only qualitative (identification) analysis of materials, but, with relevant standards, can be used for quantitative (amount) analysis. The FT-IR spectral absorbance bands of the nanoparticles of *Colpomenia sinuosa* were seen at 3435 cm^{-1} (O-H stretch, H-bonded alcohols, phenols), 2923 cm^{-1} (C-CH₃ stretch, alkanes), 2853 cm^{-1} (CH₂, alkanes), 2519 cm^{-1} (S-H stretch, thiol), 2091 cm^{-1} (C≡C stretch, alkynes), 1633 cm^{-1} (N-H bend, primary amines), 1469 cm^{-1} (C-C stretch (in-ring), aromatics), 1103 cm^{-1} (C-N stretch, aliphatic amines), 1034 cm^{-1} (C-O stretch, alcohols, ethers), 875 cm^{-1} (C-H out of plane bending, aromatics) [38], 862 cm^{-1} (C-H out of plane bending, aromatics), 712 cm^{-1} (C-Cl stretch, alkyl halides), 658 cm^{-1} (C-Br stretch, alkyl halides), 603 cm^{-1} (C-Br stretch, alkyl halides), 541 cm^{-1} (C-Br stretch, alkyl halides), and 471 cm^{-1} (S-S stretch, polysulfides). The FT-IR spectrum analysis indicates the presence of chemical bonds and the functional groups of the biomolecules responsible for the stabilization of the biosynthesized silver nanoparticles from the experimental marine brown seaweed *Colpomenia sinuosa*. The FTIR results revealed that the compounds present in the marine brown seaweed *Colpomenia sinuosa* which are aromatic, alkanes, amides or amines [39] that may act as the capping ligand in the formation of silver nanoparticles and also possible perform the stabilization of silver nanoparticles in the aqueous medium. These results obtain good agreement with the literatures [38,40].

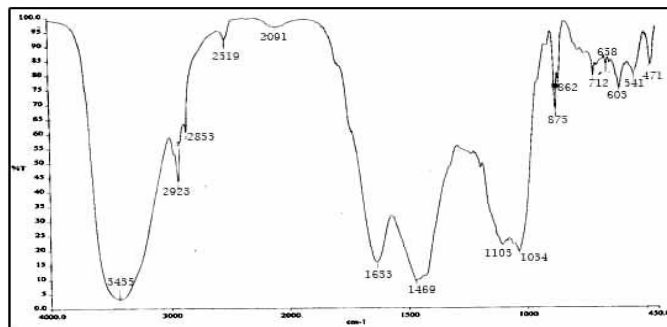


Fig. 3 FT-IR spectrum of *Colpomenia sinuosa* mediated synthesized silver nanoparticles

Table 1 FT-IR Spectral interpretation of silver nanoparticles bio-synthesized from *Colpomenia sinuosa*

Wave number (cm^{-1})	Spectral Assignments
3435	O-H stretch, H-bonded alcohols, phenols
2923	C-CH ₃ stretch, alkanes
2853	CH ₂ alkanes
2519	S-H stretch, thiol
2091	C≡C stretch, alkynes
1633	N-H bend, primary amines
1469	C-C stretch (in-ring), aromatics
1103	C-N stretch, aliphatic amines
1034	C-O stretch, alcohols, esters, ethers
875	C-H "oop", aromatics
862	C-H "oop", aromatics
712	C-Cl stretch, alkyl halides
658	C-Br stretch, alkyl halides
603	C-Br stretch, alkyl halides
541	C-Br stretch, alkyl halides
471	S-S stretch, Polysulfides

3.4 X-Ray Diffraction Pattern (XRD)

XRD is a widely used to determine the size and crystal structure of silver nanoparticles. X-ray diffractogram of the biosynthesized silver nanoparticles by the experimental seaweed *Colpomenia sinuosa* exhibits Bragg reflection corresponding to face centered cubic (fcc) type bulk silver. The broadened diffraction peaks around their base indicates that the silver nanoparticles are between nano sizes. XRD (Labtron LXR-D-A10) analysis of biosynthesized silver nanoparticles from *Colpomenia sinuosa* exhibited four distinct diffraction peaks is shown in Fig. 4. The lattice planes (1 0 0), (1 1 0), (1 1 1) and (2 0 0) were identified with the corresponding Bragg's angles of 11.58°, 32.04°, 37.89° and 46.96° respectively, which confirm the face-centered cubic structure of the silver nanoparticles. The observed peak broadening and noise were probably related to the effect of nano sized particles and the presence of various biological molecules in the reaction solution. The XRD pattern thus clearly shows that the silver nanoparticles are crystalline in nature.

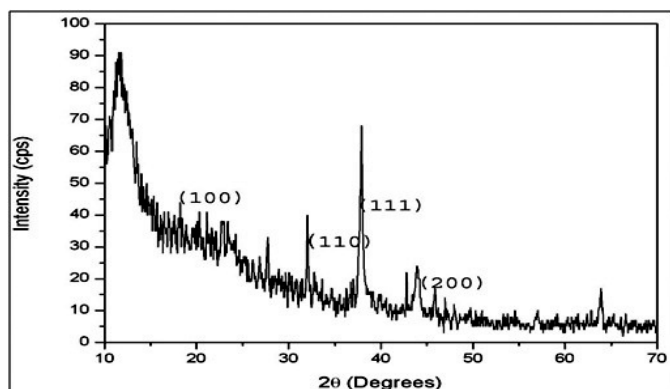


Fig. 4 X-ray diffraction analysis of silver nanoparticles biosynthesized from *Colpomenia sinuosa*

3.5 Thermo Gravimetric Analysis (TGA)

The biosynthesized silver nanoparticles from the experimental seaweed *Colpomenia sinuosa* were subjected to thermogravimetric analysis (TGA 4000 - PerkinElmer) is shown in Fig. 5. The thermogravimetric analysis was used to assess the purity and thermal stability of silver nanoparticles. The thermogram of biosynthesized silver nanoparticles from *Colpomenia sinuosa* was observed and there was no major weight loss up to 220 °C and a sharp weight loss of 12% was observed in the temperature range of 220-400 °C. TGA result shows that the purity of silver nanoparticles was 95% for biosynthesis of silver colloidal medium carried out by the ultra-sonication method [41]. There was nearly no degradation above 400 °C that accounts for the weight of the silver.

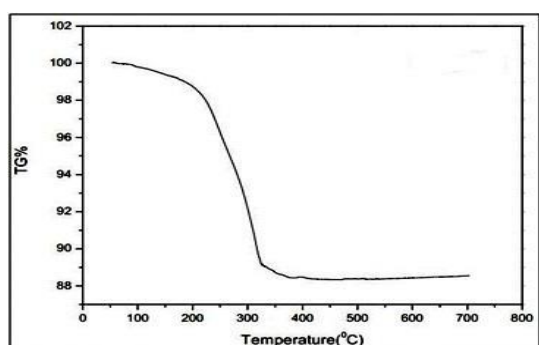


Fig. 5 TGA thermogram of silver nanoparticles bio-synthesized from *Colpomenia sinuosa*

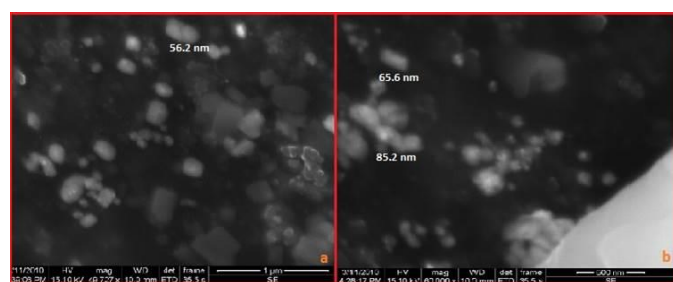


Fig. 6 Scanning Electron Micrograph of silver nanoparticles bio-synthesized from *Colpomenia sinuosa*
<https://doi.org/10.30799/jacs.219.20060101>

3.6 Scanning Electron Microscopy (SEM)

The morphology and shape of these silver nanoparticles were carried out using scanning electron microscopy (SEM Quanta - 400) is shown in Fig. 6. The silver nanoparticles synthesized from *Colpomenia sinuosa* biomass after exposure to 10^{-3} M aqueous silver nitrate solution for 2 hours showed the colloidal form of the particles in solution which micro precipitated on the surface of the biomass of the experimental seaweed. The back scattered electron image showed bright cubical and spherical area which corresponds to the silver nanoparticles indicating the structure of nanosilver. The hydrogen bond and electrostatic interactions might have played a key role along with the bioorganic capping molecules of the experimental seaweed in the formation of the morphology and shape of silver nanoparticles. The other factor in determining the shape of the silver nanoparticles may be attributed to the changes in the optical and electronic properties. The stabilization of the nanoparticles by the capping agent might be due to nanoparticles were not in direct contact within the aggregates. Thus, the shape and morphology of the biosynthesized nanoparticles by the experimental brown seaweed *Colpomenia sinuosa* was clearly revealed by the SEM results.

3.7 Transmission Electron Microscopy (TEM)

Further insight on morphology and the size details of the biosynthesized silver nanoparticles by the experimental seaweed *Colpomenia sinuosa* were investigated using high resolution transmission electron microscopy (HR-TEM JEOL 3010) is shown in Fig. 7. The HR-TEM images of silver nanoparticles clearly revealed the formation of spherical and hexagonal shaped nanoparticles. The majority of the nanoparticles appeared spherical and only a small percentage was elongated particles that ranged in size from 5 to 50 nm. The average means size of silver nanoparticles was 34 nm.

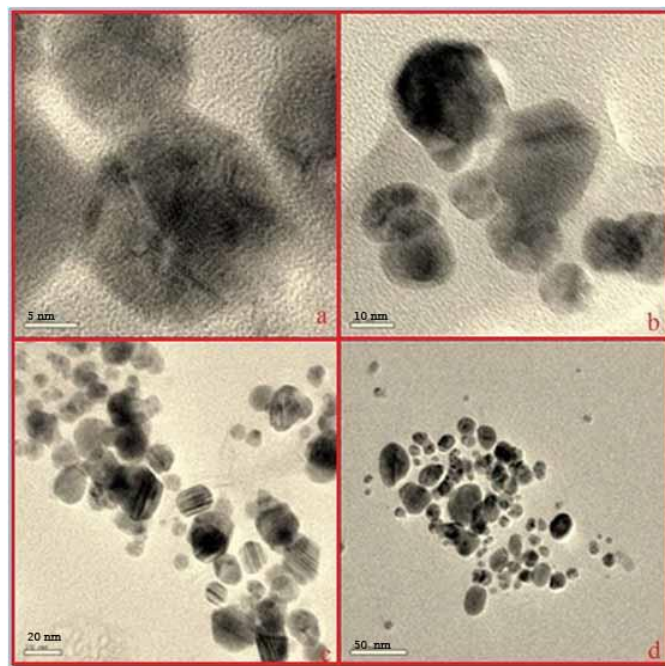


Fig. 7 HR-TEM images of silver nanoparticles bio-synthesized from *Colpomenia sinuosa*

4. Conclusion

In this work silver nanoparticles were synthesized by the reduction of silver nitrate using the extract of marine brown seaweed *Colpomenia sinuosa*. The reaction mixture was successfully optimized to increase the yield of silver nanoparticles production using UV-Vis analysis results. The optimum conditions were as follows: 500 g extract concentration, 10^{-3} M aqueous AgNO_3 solution, 37 °C and pH 5.09. The *Colpomenia sinuosa* seaweed extract acts as both reducing and stabilizing agents for the synthesis of silver nanoparticles, which were confirmed by FTIR. SEM analysis revealed that the silver nanoparticles ranged between 54 nm to 65 nm in size and were spherical with uniform distribution. TEM analysis revealed nearly spherical and hexagonal structures of the silver nanoparticles. The XRD pattern of silver nanoparticles showed a face-centered cubic crystal structure. The purity and thermal stability of silver nanoparticles were detected by TGA analysis, which was closely related to that of bulk metallic silver, which indicates its purity. The benefits of using

seaweeds via green synthesis for bio synthesis of silver nanoparticles is an environmental-friendly process that promotes energy efficiency, cost effectiveness, and protects human health while leads to inherently safer products and lesser waste. This environmental-friendly method of synthesizing silver nanoparticles as alternative measure to conventional physical and chemical methods has an enormous potential to be used in low cost production applications.

Acknowledgement

This work was carried out in part through the use of the scanning electron microscopy (SEM) and transmission electron microscopy (TEM) at Sophisticated Analysis Instrument Facility (SAIF), Indian Institute of Technology (IIT) – Madras, Tamilnadu, India. The thermogravimetric analysis (TGA) was carried out at Department of Chemistry, Anna University, Chennai, Tamilnadu, India. The authors thank the above institutions for their instrumentation facility for the completion of the work. We also sincerely thank Mr. Raju sea diver who helped me in the collection of the sea weeds from south east coast of Tamilnadu, India.

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